

Australian eggs

Purpose/Scope: This SOP provides a method for conducting *Salmonella* sampling in deep litter sheds with no pens or partition



MATERIALS NEEDED

- Cotton guaze swabs, can use either:
 - See instructions on how to make your own*or,
 - Tampons or,
 - Supplied by laboratory
- 1.5m cotton string
- Disposable latex gloves
- Sample transport liquid (peptone water)
- *Whirl-Pak® bags or screw top plastic jar
- Scissors
- Permanent marker
- Laboratory sample submission form
- Plastic post satchel for transporting swabs to the laboratory
- Plastic container for swabbed samples
- * Making cotton gauze swabs
- ^ https://www.whirl-pak.com/ whirl-pak-bags-general-information

MAKING THE COTTON GAUZE SWABS

1 Obtain a 10cm x 10cm cotton gauze and fold onto itself in a pleated pattern.



Figure 1 Image: Michael J et al. 2020

2 Continue folding gauze to form a pad.



Figure 2 Image: Michael J et al. 2020

3 Tie the cotton string around the centre of the cotton gauze.



Figure 3 Image: Michael J et al. 2020

4 Wind string around the cotton gauze.



Figure 4

- 5 Place the required number of swabs for each shed into their own plastic container or Whirl-Pak® bag.
- 6 Store the rest in a dry, secure place.

PROCEDURE

Step 1

Get prepared

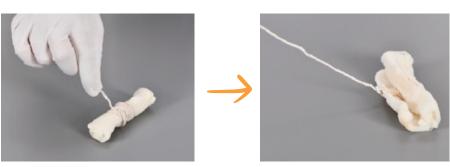
- 1 Notify the laboratory 24 hours in advance of sending the swab samples.
- Obtain a sample submission form from the laboratory.
- 3 Prepare **three (3) swabs** per shed.

Step 2

Swab the shed

- 1 Wash your hands.
- 2 Put on a pair of disposable latex gloves.
- 3 Moisten **Swab 1** with water from the drinkers or solution provided by the laboratory.
- 4 Hold **Swab 1** by the string and unravel (Figure 5).

Figure 5. Hold the swab by the string and unravel (Romer Labs)



5 Drag **Swab 1** twice the full length of the shed over different areas of litter in the patterns described in Figure 6.

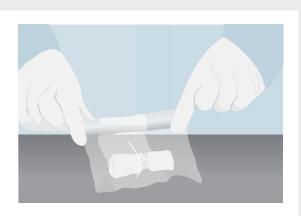


Swab 1 should be considered finished when the swab is back to where it started from (Figure 6).

PROCEDURE

- 7 The string should not be included in the sample sent to the laboratory, cut the string from **Swab 1** with a pair of scissors.
- 8 Place **Swab 1** in a Whirl-Pak® bag or screw top plastic jar (Figure 7).

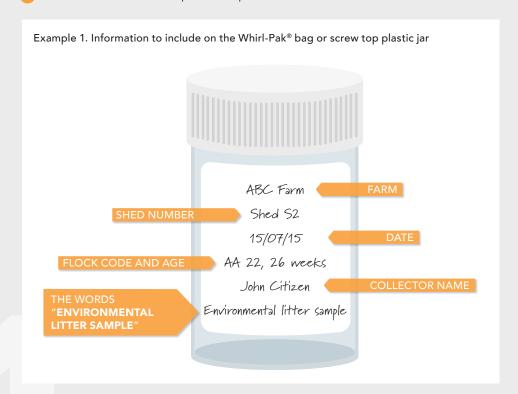
Figure 7. Put swab into Whirl-Pak® bag (Romer Labs)



- Seal the bag or plastic jar.
- 10 Repeat procedure 3 to 9 with Swab 2 and Swab 3, using one Whirl-Pak® bag or plastic jar per swab. If gloves come into contact with litter or manure they should be changed between swabs.

Step 3Pack the samples

- 1 Each sample should be placed in it's own Whirl-Pak® bag or screw top plastic jar. Clearly label each bag or jar with permanent marker.
- 2 Include information as per Example 1.



3 Complete the laboratory sample submission form (always record on submission sheets as "ENVIRONMENTAL LITTER SAMPLES").

PROCEDURE

Step 4Submit the samples

1 Pack the swabs that are in the bags (Figure 8A) securely into a plastic container (Figure 8B) and put the container into a plastic post satchel (Figure 8C).

ABC Farm
Shed 52
15/07/15
AA 22, 26 weeks
John Citizen
Environmental litter
sample

3B



https://ie.vwr.com/store/ product/17962031/samplecontainer-with-screw-capsterilin#gallery-1



https://auspost.com.au/shop/ product/flat-rate-smallsatchel-10-pack-059049131?fm =recommendations:shop:1

- 2 Put the completed sample submission form into the same plastic post satchel as the swabs.
- 3 Post the samples to the diagnostic laboratory.
- 4 If the swabs cannot be posted on the same day, store the swabs in the fridge (between 4 and 8°C) until ready to be posted. Conduct procedures 1 to 4 as soon as possible.

Swabs must not be frozen.

REFERENCE

Michael J. Sikorski, Myron M. Levine 2020 Reviving the "Moore Swab": A Classic Environmental Surveillance Tool Involving Filtration of Flowing Surface Water and Sewage Water To Recover Typhoidal *Salmonella* Bacteria

Applied and Environmental Microbiology, 86 (13) e00060-20; **DOI:** 10.1128/AEM.00060-20)

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